

516 Rec'd PCT/PTO 23 JUL 1999

2FORM PTO-1390
OFFICE
(REV. 1094)

U.S. DEPARTMENT OF COMMERCE PATENT AND TRADEMARK

ATTORNEY'S DOCKET NUMBER
C1041/7005

U.S. APPLICATION NO. (If known, see 37 CFR 1.5)

TRANSMITTAL LETTER TO THE UNITED STATES
DESIGNATED/ELECTED OFFICE (DO/EO/US)
CONCERNING A FILING UNDER 35 U.S.C. 371

09/355254

INTERNATIONAL APPLICATION NO.
PCT/EP98/00367INTERNATIONAL FILING DATE
23 January 1998 (23.01.98)PRIORITY DATE CLAIMED
23 January 1997 (23.01.97)TITLE OF INVENTION
PHARMACEUTICAL COMPOSITIONS COMPRISING A POLYNUCLEOTIDE AND OPTIONALLY AN ANTIGEN ESPECIALLY
FOR VACCINATIONAPPLICANT(S) FOR DO/EO/US
WAGNER, Hermann, LIPFORD, Grayson, B., HEEG, Klaus

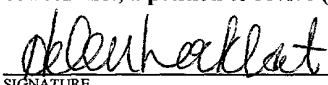
Applicant herewith submits to the United States Designated/Elected Office (DO/EO/US) the following items and other information:

1. ☒ This is a **FIRST** submission of items concerning a filing under 35 U.S.C. 371.
2. ☐ This is a **SECOND** or **SUBSEQUENT** submission of items concerning a filing under 35 U.S.C. 371.
3. ☒ This express request to begin national procedures (35 U.S.C. 371(f) at any time rather than delay examination until the expiration of the applicable time limit set in 35 U.S.C. 371(b) and PCT Articles 22 and 39(1).
4. ☒ A proper Demand for International Preliminary Examination was made by the 19th month from the earliest claimed priority date.
5. ☒ A copy of the International Application as filed (35 U.S.C. 371(c)(2)).
 - a. ☐ is transmitted herewith (required only if not transmitted by the International Bureau).
 - b. ☒ has been transmitted by the International Bureau.
 - c. ☐ is not required, as the application was filed in the United States Receiving Office (RO/US).
6. ☐ A translation of the International Application into English (35 U.S.C. 371(c)(2)) with verification of translation.
7. ☒ Amendments to the claims of the International Application under PCT Article 19 (35 U.S.C. 371(c)(3)).
 - a. ☐ are transmitted herewith (required only if not transmitted by the International Bureau).
 - b. ☐ have been transmitted by the International Bureau.
 - c. ☐ have not been made; however, the time limit for making such amendments has NOT expired.
 - d. ☒ have not been made and will not be made.
8. ☐ A translation of the amendments to the claims under PCT Article 19 (35 U.S.C. 371(c)(3)).
9. ☐ An oath or declaration of the inventor(s) (35 U.S.C. 371(c)(4)).
10. ☐ A translation of the annexes to the International Preliminary Examination Report under PCT Article 36 (35 U.S.C. 371(C)(5)).

Items 11. To 16. Below concern document(s) or information included:

11. ☒ An Information Disclosure Statement under 37 CFR 1.97 and 1.98 with references.
12. ☐ An assignment document for recording. A separate cover sheet in compliance with 37 CFR 3.28 and 3.31 is included.
13. ☐ A FIRST preliminary amendment.
☐ A SECOND or SUBSEQUENT preliminary amendment.
14. ☐ A substitute specification (submitted as a first Preliminary Amendment).
15. ☐ A change of power of attorney and/or address letter.
16. ☒ Other items or information:
 - Notification Concerning Informal Communications With the Applicant; International Preliminary Examination Report; Copy of Response to EPO Examiner Halle submitting requested Amended Claims in which the SEQ ID Nos. and sequences have been adapted
 - PCT Request Form PCT/RO/101
 - Nucleotide and/or Amino Acid Sequence Listing
 - Statement Under 37 C.F.R. 1.821(f) with diskette
 - Mailed via Express Mailing Label No. EL310412446US
 - Post Card

Express Mail Label No. EL310412446US

U.S. APPLICATION NO (If known, see 37 CFR 1.5) 09/355254		INTERNATIONAL APPLICATION PCT/EP98/00367		ATTORNEY'S DOCKET NUMBER C1041/7005	
17. X The following fees are submitted:				CALCULATIONS PTO USE ONLY	
BASIC NATIONAL FEE (37 CFR 1.492(a)(1)-(5)): Search Report has been prepared by the EPO or JPO \$840.00 International preliminary examination fee paid to USPTO (37 CFR 1.482) \$670.00 No international preliminary examination fee paid to USPTO (37 CFR 1.482) but international search fee paid to USPTO (37 CFR 1.445(a)(2)).. \$760.00 Neither international preliminary examination fee (37 CFR 1.482) nor international search fee (37 CFR 1.445(a)(2)) paid to USPTO..... \$970.00 International preliminary examination fee paid to USPTO (37 CFR 1.482) and all claims satisfied provisions of PCT Article 33(2)-(4) \$96.00 ENTER APPROPRIATE BASIC FEE AMOUNT =				\$ 840.00	
Surcharge of \$130.00 for furnishing the oath or declaration later than <input type="checkbox"/> 20 X 30 months from the earliest claimed priority date (37 CFR 1.492(e)).					
CLAIMS	NUMBER FILED	NUMBER EXTRA	RATE		
Total Claims	12 - 20 =	0	X \$18.00	\$	
Independent Claims	5 - 3 =	2	X \$78.00	\$ 156.00	
MULTIPLE DEPENDENT CLAIM(S) (if applicable)			+\$260.00	\$	
TOTAL OF ABOVE CALCULATIONS =				\$ 996.00	
Reduction by 1/2 for filing by small entity, if applicable. Verified Small Entity Statement must also be filed (Note 37 CFR 1.9, 1.27, 1.28).				\$ 498.00	
SUBTOTAL =				\$ 498.00	
Processing fee of \$130.00 for furnishing the English translation later than <input type="checkbox"/> 20 <input type="checkbox"/> 30 months from the earliest claimed priority date (37 CFR 1.492(f)).				\$	
TOTAL NATIONAL FEE =				\$	
Fee for recording the enclosed assignment (37 CFR 1.21(h)). The assignment must be accompanied by an appropriate coversheet (37 CFR 3.28, 3.31). \$40.00 per property +				\$	
TOTAL FEES ENCLOSED =				\$ 498.00	
				Amount to be: refunded	\$
				charged	\$
a. A check in the amount of \$_____ to cover the above fees is enclosed. b. X Please charge by Deposit Account No. <u>23/2825</u> In the amount of \$ <u>498.00</u> To cover the above fees. c. X The commissioner is hereby authorized to charge any additional fees which may be required, or credit any overpayment to Deposit Account No. 23/2825. A duplicate of this sheet is enclosed.					
NOTE: Where an appropriate time limit under 37 CFR 1.494 or 1.495 has not been met, a petition to revive (37 CFR 1.137(a) or (b)) must be filed and granted to restore the application to pending status.					
SEND ALL CORRESPONDENCE TO				 SIGNATURE	
Helen C. Lockhart WOLF, GREENFIELD & SACKS, P.C. 600 Atlantic Avenue Boston, Massachusetts 02210				<u>Helen C. Lockhart</u> NAME	
				<u>39,248</u> REGISTRATION NO	

**Pharmaceutical composition comprising a polynucleotide
and optionally an antigen especially for vaccination**

The present invention relates to pharmaceutical compositions comprising at least one fragment of a polynucleotide, preferably at least one antigen, and optionally a pharmaceutically acceptable carrier and/or diluent. In accordance with the present invention was found that the introduction of the pharmaceutical composition into vertebrates will achieve regulation of growth, induction of cellular transcription and translation, protein synthesis, protein expression or protein secretion. The pharmaceutical compositions are useful in vaccination protocols but also in any other therapeutic situation in which immunomodulation is of benefit, such as sub-optimal immune responses, reaction to pathogens, tolerance or autoimmunity.

It is known that cells of the immune system are exported from the bone marrow and undergo a series of differentiation events which confer upon them the capacity to recognize and control foreign pathogens and cancer cells by discriminating between self versus non-self. These differentiation and education events are tightly controlled by cell surface receptor engagement via intracellular signal transduction and the milieu of autocrine, paracrine and endocrine soluble ligands, typically referred to as cytokines. Cell to cell interaction occurs in discrete locations such as the thymus, spleen or lymph nodes but also in the periphery. The system thus balances receptor and cytokine input signals to regulate cellular proliferation, differentiation and maturation of immune effector cells [Paul, Cell, 57:521 (1989)]. Through outside intervention the immune system can be manipulated, namely enhanced, e.g. by vaccination or cytokine therapies, or suppressed, e.g. by drug intervention or cytokine therapies.

The immune system of vertebrates consists of several interacting components. The best characterized and most important parts are the humoral and cellular (cytolytic) branches. Humoral immunity involves antibodies, proteins which are secreted into the body fluids and which directly recognize an antigen. The cellular system, in contrast, relies on special cells which recognize and kill other cells which are producing foreign antigens. This basic functional division reflects two different strategies of the immune defense. Humoral immunity is mainly directed at antigens which are exogenous to the host's somatic cells or on the surface of cells whereas the cellular system responds to antigens which are actively synthesized within cells or derived from phagocytosed exogenous antigens.

Antibody molecules, the effectors of humoral immunity, are secreted by special B lymphoid cells, B cells, in response to antigen, co-receptor stimulation and cytokines. Antibodies can bind to and inactivate antigen directly (neutralizing antibodies) or activate other cells of the immune system to destroy the antigen depending on isotype; IgM, IgG1, etc. Isotype class switching in B cells is controlled, inter alia, by cytokine milieu. Abnormalities in antigen response, co-receptor engagement or cytokine milieu can lead to suboptimal immune responses, tolerance or autoimmunity.

Cellular immune recognition is mediated by a special class of lymphoid cells, the T cells. These cells do not recognize whole antigens but instead respond to degraded peptide fragments thereof which appear on the surface of the antigen presenting cells bound to surface proteins called major histocompatibility complex (MHC) molecules. Two subgroups of T cells exist: the CD4 T cells recognize peptide fragments bound to MHC class II molecules while CD8 T cells recognize peptide fragments bound to MHC class I molecules. CD8 T cells include the population of cytotoxic T cells (CTL) able to specifically lyse antigen presenting cells. Essentially all nucleated cells have class I molecules. It is believed that proteins produced within the cell are continually degraded to peptides as part of normal cellular metabolism. These fragments are bound to the MHC molecules and are transported to the cell surface. Thus, the cellular immune system is constantly monitoring the spectra of proteins produced in all cells in the body

and is poised to eliminate any cells producing foreign antigens or abnormal self antigens. CD4 T cells recognize mainly exogenous antigens that were taken up by antigen processing cells wherein the antigen is degraded and appears as a peptide fragment on class II MHC molecules. The effector function of CD4 cells is primarily regulation of immune responses by release of cytokines. According to the cytokine profiles secreted, two subclasses of CD4 cells have been defined, the TH1 and TH2 cells. It is believed that in various infections and allergic and autoimmune diseases the type of the CD4 T cell subclass activated (TH1 vs. TH2) critically influences the outcome of the immune response.

Vaccination is the process of preparing a human or an animal to respond to an antigen. Vaccination is more complex than immune recognition and involves not only B cells and cytotoxic T cells but other types of lymphoid cells as well. During vaccination, cells which recognize the antigen (B cells or T cells) are clonally expanded. In addition, the population of ancillary cells (helper T cells which provide co-receptor and cytokine stimulation) specific for the antigen also increase. Vaccination also involves specialized antigen presenting cells which can process the antigen and display it in a form which can stimulate one of the two pathways (macrophages and dendritic cells).

A foreign antigen is introduced into a human or an animal where it activates specific B cells by binding to surface immunoglobulins. It is also taken up by antigen processing cells, wherein it is degraded, and appears in fragments on the surface of these cells bound to Class II MHC molecules. Peptides bound to class II molecules are capable of stimulating the helper class of T cells (CD4 T cells). Both helper T cells and activated B cells are required to produce active humoral immunization. Cellular immunity is stimulated by a similar mechanism but entry into the MHC I presentation pathway of antigen presenting cells is typically by intracellular pathogen replication and not achieved by injection of protein antigen only.

Standard vaccination schemes nearly always produce a humoral immune response. The humoral system protects a vaccinated individual from subsequent challenge from a

pathogen and can prevent the spread of an intracellular infection if the pathogen goes through an extracellular phase during its life cycle; however, it can do relatively little to eliminate intracellular pathogens. Cytotoxic immunity complements the humoral system by eliminating the infected cells and cancer cells. Thus, effective vaccination should advantageously activate both types of immunity.

A cytotoxic T cell response is necessary to remove intracellular pathogens such as viruses as well as malignant cells. It has proven difficult to present an exogenously administered antigen in adequate concentrations in conjunction with Class I molecules to assure an adequate response. This has severely hindered the development of vaccines against tumor-specific antigens (e.g., on breast or colon cancer cells), and against weakly immunogenic viral proteins (e.g., HIV, Herpes, non-A, non-B hepatitis, CMV and EBV). It would be desirable to provide a cellular immune response alone in immunizing against agents such as viruses for which antibodies have been shown to enhance infectivity. It would also be useful to provide such a response against both chronic and latent viral infections and against malignant cells.

Directed and elevated immune responses to antigens can be achieved by the use of adjuvants and/or delivery vehicles. The term "immune adjuvant" refers to compounds which when administered to an individual or tested in vitro, increase the immune response to an antigen. Some antigens are weakly immunogenic when administered alone or are toxic at the concentration which evokes immune responses. An immune adjuvant may enhance the immune response of the individual to the antigen by making the antigen more immunogenic. The adjuvant effect may also lower the dose of antigen necessary to achieve an immune response by enhancing presentation, influence the cytokine milieu or alter co-receptor expression on antigen presenting cells.

Recently, it has been demonstrated that oligonucleotides and oligonucleotide derivatives may have a significant impact on certain biological processes. Thus, it has been shown that phosphorothioate oligonucleotides may have an influence on the regulation of gene expression [Bielinska et al., Science, vol. 250 (1990), p. 997-250].

Krieg et al. [Nature, vol. 374 (1995), p. 546-549] report that bacterial DNA may trigger direct B cell activation. They disclose that bacterial DNA and synthetic oligodeoxynucleotides containing unmethylated CpG dinucleotides induce murine B cells to proliferate and secrete immunoglobulin in vitro and in vivo. Yet, the system developed by Krieg et al. has not proven successful in every instance to trigger, modulate or enhance an immune response that is beneficial to the patient. In some instances, the system developed by Krieg even showed adverse effects. Thus, the technical problem underlying the present invention was to provide a means that is effectively and beneficially applicable in a wide variety of situations where stimulation, modulation or triggering of an immunological reaction is desired.

The solution to said technical problem is activated by providing the embodiments characterized in the claims.

Accordingly, the present invention relates to a pharmaceutical composition comprising at least one fragment of a polynucleotide and at least one antigen and, optionally, a pharmaceutically acceptable carrier and/or diluent.

The term "polynucleotide" in the sense of the present invention comprises all types of polynucleotides as well as derivatives thereof, e.g. RNA, PNA or DNA whereby, however, DNA polynucleotides are preferred. The term "fragment of a polynucleotide", as used in accordance with the present invention, may relate to a fragment generated from a polynucleotide or to a nucleic acid molecule that is shorter than a polynucleotide, such as an oligonucleotide which may be of synthetic origin.

The term "antigen" in the sense of the present invention means a molecule that can elicit an immune response. The immune response may be either humoral, i.e. antibody-mediated, or cellular, i.e. cell-mediated. An antigen that evokes an immune response is commonly referred to as immunogen. Generally only foreign or "non-self" molecules are immunogenic. It should be understood, however, that in the sense of the present invention, the term antigen comprises also certain "self" molecules such as tumor cells,

tumor markers or self antigens in autoimmunity. Those compounds may not be foreign to the host to be treated, but may be comprised under the term antigen, since sometimes an immune response to self molecules is desired. In that case exogenous addition of antigen to the pharmaceutical composition may not be required. The fragment of the polynucleotide as well as the antigen may be of natural, synthetic or semisynthetic origin. Particularly, in the case that the antigen is of natural origin, it may be processed prior to administration.

Usually the larger and more complex a molecule is, the more immunogenic sites it will have. A single antigen may contain many epitopes which are specific areas of the molecule with a three-dimensional configuration that induces an immune response. Complex molecules, such as large proteins composed of many different amino acids contain more epitopes than a comparatively simple polysaccharide composed of two or three monosaccharide repeats. The immune response to a given antigen can vary greatly among species and individuals within a species due to immune regulation genes. The pharmaceutical compositions of the present invention comprise therefore specific polynucleotides which provoke a suitable immune response.

It is preferred that the fragment of the polynucleotide as used in the present invention comprises the sequence of a binding site for transcription factors or parts thereof or that the sequence of the polynucleotide is complementary to said binding site for transcription factors or a part thereof.

The expression of individual genes is a rather complex process. These processes are mediated by several specific regulatory DNA regions found in the promotor regions of almost all genes. These regulatory sequences are frequently referred to as response elements. They are binding sites for sequence-specific DNA binding proteins which are called transcription factors. Some transcription factors are general purpose factors (basal transcription factors) required for transcription of all genes while others act on specific genes or classes of genes by binding in a sequence-specific manner to response elements and other sequence motives within the corresponding gene

promoters. The expression of many of these transcription factors is developmentally and also tissue-specifically controlled and is itself subject to the action of other transcription factors and other accessory proteins such as nuclear receptors. Binding sites for transcription factors are often clustered and a variety of transcription factors have been found to form complexes with others or to compete with others for binding to overlapping DNA-binding motives. Several structural motives have been found within those regions of transcription factor proteins recognizing and contacting DNA. Within each of these structural motives there are often families of related proteins that recognize similar DNA sequences and are conserved throughout the eukaryotic kingdom.

In the course of the present invention it has been surprisingly found that bacterial sequences as described in the prior art may cause severe side effects such as lethal shock. It has been found, however, that other DNA sequences may have a beneficial effect on mammals and may therefore be used for the preparation of pharmaceutical compositions having a beneficial effect on the immune system. Thus, selection and identification of such beneficial DNA sequences may be effected on the basis of the teachings of the present invention.

The present invention also relates to a pharmaceutical composition comprising

- (a) a polynucleotide or an oligonucleotide comprising the sequence of a binding site for transcription factors or a part thereof or a polynucleotide or an oligonucleotide comprising a sequence which is complementary to said binding site for transcription factors or a part thereof; and optionally
- (b) a pharmaceutically acceptable carrier and/or diluent.

In the course of the present invention it has been found that polynucleotides are preferred which have a sequence corresponding to the binding site of transcription factors or which are complementary thereto. Generally it is sufficient that at least a part of said binding site is contained within the polynucleotides. This embodiment of the

invention is advantageously employed in conjunction with the above cited "self" molecules.

Advantageously, the pharmaceutical compositions of the present invention comprise inexpensive, stable and safe immune adjuvants and immunomodulatory substances for human and animal prophylactic and therapeutic use.

As has been stated above, the present invention relates to a pharmaceutical composition comprising at least one of the aforementioned regulatory sequences or antigens, either alone or in combination, and optionally a pharmaceutically acceptable carrier or excipient. Examples of suitable pharmaceutical carriers are well known in the art and include phosphate buffered saline solutions, water, emulsions, such as oil/water emulsions, various types of wetting agents, sterile solutions etc. Compositions comprising such carriers can be formulated by well known conventional methods. These pharmaceutical compositions can be administered to the subject at a suitable dose. Administration of the suitable compositions may be effected by different ways, e.g., by intravenous, intraperitoneal, subcutaneous, intramuscular, topical or intradermal administration. The dosage regimen will be determined by the attending physician and other clinical factors. As is well known in the medical arts, dosages for any one patient depends upon many factors, including the patient's size, body surface area, age, the particular compound to be administered, sex, time and route of administration, general health, and other drugs being administered concurrently. Generally, the regimen as a regular administration of the pharmaceutical composition should be in the range of 1 μ g to 10 mg units per day. If the regimen is a continuous infusion, it should also be in the range of 1 μ g to 10 mg units per kilogram of body weight per minute, respectively. Progress can be monitored by periodic assessment. Dosages will vary but a preferred dosage for intravenous administration of DNA is from approximately 10^6 to 10^{22} copies of the DNA molecule. The compositions of the invention may be administered locally or systemically. Administration will generally be parenterally, e.g., intravenously; DNA may also be administered directly to the target site, e.g., by biolistic delivery to an internal or external target site or by catheter to a site in an artery.

Advantageously, in the pharmaceutical compositions of the present invention liposomes can serve as carriers for the direction of antigen to antigen presenting cells. It has been demonstrated that liposomes can serve to heighten some humoral immune responses and to provide for exogeneously administered antigen a vehicle for entry into the MHC class I presentation pathway thus allowing the stimulation of cytotoxic T cells.

It is envisaged by the present invention that the various DNA oligonucleotides and/or polynucleotides or fragments thereof are administered either alone or in any combination using standard vectors and/or gene delivery systems, and optionally together with an appropriate compound, and/or together with a pharmaceutically acceptable carrier or excipient. Subsequent to administration, said DNA oligonucleotides and/or polynucleotides may be stably integrated into the genome of the mammal. On the other hand, viral vectors may be used which are specific for certain cells or tissues and persist therein. Suitable pharmaceutical carriers and excipients are well known in the art. The pharmaceutical compositions prepared according to the invention can be used for the prevention or treatment or delaying of different kinds of diseases, which are related to the expression or overexpression of genes.

Furthermore, it is possible to use a pharmaceutical composition of the invention which comprises DNA oligonucleotide in gene therapy. Suitable gene delivery systems may include liposomes, receptor-mediated delivery systems, naked DNA, and viral vectors such as herpes viruses, retroviruses, adenoviruses, and adeno-associated viruses, among others. Delivery of nucleic acids to a specific site in the body for gene therapy or antisense therapy may also be accomplished using a biolistic delivery system, such as that described by Williams (Proc. Natl. Acad. Sci. USA 88 (1991), 2726-2729).

Standard methods for transfecting cells with recombinant DNA are well known to those skilled in the art of molecular biology, see, e.g., WO 94/29469. Gene therapy and antisense therapy to diseases mentioned in accordance with the invention may be carried out by directly administering the DNA oligonucleotide to a patient or by

transfecting cells with the DNA oligonucleotide *ex vivo* and infusing the transfected cells into the patient. Furthermore, research pertaining to gene transfer into cells of the germ line is one of the fastest growing fields in reproductive biology. Gene therapy, which is based on introducing therapeutic genes into cells by *ex-vivo* or *in-vivo* techniques is one of the most important applications of gene transfer. Suitable vectors and methods for *in-vitro* or *in-vivo* gene therapy are described in the literature and are known to the person skilled in the art; see, e.g., WO94/29469, WO 97/00957 or Schaper (Current Opinion in Biotechnology 7 (1996), 635-640) and references cited therein. The DNA molecules and vectors comprised in the pharmaceutical composition of the invention may be designed for direct introduction or for introduction via liposomes, or viral vectors (e.g. adenoviral, retroviral) containing said recombinant DNA molecule into the cell. Preferably, said cell is a germ line cell, embryonic cell, or egg cell or derived therefrom. The pharmaceutical compositions according to the invention can be used for the treatment of diseases hitherto unknown as being related to susceptible to cytokine presence or concentration. An embryonic cell can be for example an embryonic stem cell as described in, e.g., Nagy, Proc. Natl. Acad. Sci. 90 (1993) 8424-8428.

In a preferred embodiment of the present invention the oligonucleotide or polynucleotide is a DNA oligonucleotide. DNA oligonucleotides as used in the present invention are preferably short fragments of a DNA having about 100, preferably five to about 40 and most preferably 15 to about 25 nucleotides.

The DNA polynucleotide fragments which are preferably oligomers provide when applied in relatively low quantities in the range of 0.1 to 10 μ g the effect of an immune adjuvant and immunomodulatory substance together with low toxicity and low side effects. For example can a combination of a suitable DNA oligomer and tumor cells or specific tumor markers induce tumor regression. The invention also relates to corresponding uses.

In a preferred embodiment of the present invention the DNA oligonucleotide is single-stranded or double-stranded, whereby single-stranded DNA is especially preferred.

In a further preferred embodiment of the present invention, said sequences of binding sites for transcription factors are binding sites of transcription factors of cytokines.

In another preferred embodiment of the present invention, the polynucleotide contained in said pharmaceutical composition comprises the sequence 5'PuPuCGPyC or a non-toxic derivative thereof wherein Pu means purine and comprises adenine and guanine and Py has the meaning of pyrimidine and comprises cytosine, thymine and uracil and wherein A means adenine, C means cytosine and G means guanine.

Non-toxicity of said derivatives can be effected according to conventional protocols.

Those preferred sequences of the present invention differ slightly but biologically significantly from the CpG motives as disclosed in the prior art.

It is furthermore preferred that in the pharmaceutical composition of the invention, said binding site is or is derived from a eukaryotic binding site.

The term "derived from" is intended to mean in accordance with this invention that one or more nucleotides from a naturally occurring binding site are changed. Said change can be effected according to conventional protocols such as mutagenesis protocols.

It is particularly preferred that said eukaryotic binding site is a binding site for a cytokine.

As regards the part of said binding site for said transcription factor or its complementary sequence, it is preferred that said part is a motif or a complementary sequence thereof. Motives of transcription factors are well known in the art and need not be discussed here any further.

It is furthermore particularly advantageous that said part comprises at least 7 nucleotides. Whereas this length of said part (and, of course, of the complete

oligonucleotide as well) has been particularly advantageous, it may also comprise significantly more than 7 nucleotides.

It is preferred that the polynucleotide comprises at least one phosphorothioate linkage. In the preferred phosphorothioate derivatives of the polynucleotides at least one oxygen atom of the phosphate backbone of the polynucleotide is replaced by a sulphur atom. Those compounds are more stable against degradation.

It is also possible to add to the pharmaceutical composition of the present invention further classical adjuvants which are known to the person skilled in the art. Examples may be preparations from the cell walls of bacteria. The pharmaceutical compositions of the present invention advantageously comprise also additives, otherwise known as pharmaceutically acceptable carriers and/or diluents known to the person skilled in the art depending on the administrative way e.g. oral, parenteral or rectal.

It is possible to use in accordance with the present invention a wide variety of antigens. Preferred antigens are selected from the group comprising peptides, polypeptides, steroides and tumor cells.

Further examples for preferred antigens may be killed intact bacteria, toxoides (i.e. toxins that are still immunogenic but are rendered biologically inactive by treatment with a chemical, heat or mutation), subunit vaccines in which only the non-toxic portion of the molecule is used or life-attenuated vaccines in which a viral or bacterial strain is rendered non-pathogenic (e.g. by passaging the virus in cell culture or deletion of bacterial genes), but is still able to multiply to a limited degree thereby eliciting protective immune response in the absence of disease symptoms.

Since the pharmaceutical composition of the present invention can also be used for the treatment and/or prophylaxis of such diseases which are not caused by foreign organisms the antigen may be also an antigen of the own body like a tumor antigen. For the treatment of autoimmune diseases or in order to positively influence the tolerance it

may also be effective to use antigens derived from the body to be treated. In that cases it may not be required to add the antigen to the pharmaceutical composition since the antigen is already present in the host.

In the accomplishment of the foregoing objectives of the invention, vaccine formulations are made which induce both humoral and cellular immune responses to antigen using preferably sequences with non-toxic embodiments of the motif 5'Pu-Pu-CpG-Py-Py-3'. Some sequences, especially with the motif 5'Pu-Pu-CpG-Py-Py-3' can be toxic and lethal. It is therefore another aspect of the invention to modify these sequences in such a way that immune adjuvant activity is maintained but toxicity is eliminated. In addition, DNA sequences are described which do not follow this motif but serve as immune adjuvant and immune response modifiers. A common aspect of the invention is that sequences from eukaryotic promoters are used. ssDNA sequences containing palindromic and non-palindromic transcription response elements, that is sequences recognized by transcription factors (proteins which regulate gene transcription) serve also as immune adjuvant or immune response modifiers in accordance with the invention. These sequences are capable of modulating lymphocyte cell surface markers and cytokine release in vitro and in vivo. In yet another aspect of the invention, the methods of treatment are pharmaceutical compositions and appropriate utilized therapeutic approaches for treating immune system tolerance and control of tumor.

The teaching of the present invention can be used for modulating the immune response to antigen by using certain DNA oligomers. The particular DNA depends on the desired outcome. Its sequence can be determined by the person skilled in the art without further ado on the basis of the teachings of the present invention. The invention works in vitro and in vivo in warm blooded animals.

A common aspect of the invention is to use the polynucleotide fragment and, in particular, certain DNA oligonucleotides to specifically influence the regulation and signalling machinery of eukaryotic cells. In particular, growth, induction of cellular transcription and translation, protein synthesis or protein secretion can be modified by

The invention concerns also the use of specific oligonucleotides or polynucleotide fragments as defined herein above which are preferably DNA for the preparation of a pharmaceutical composition which have the effect to interfere, to modulate and to regulate responses of the innate and acquired immune system. Those include enhancement of immune responses (including vaccination), modulation of immune responses and suppression of immune responses.

Use of DNA oligos to modulate responses of the acquired immune system is also an aspect of the present invention. Desired immune responses can be stimulated while

adverse effects can be suppressed by DNA oligonucleotides. DNA oligonucleotides can shift an immune response to a TH1-type or a TH2-type of reactivity. This effect modulates the response during parasitic infections (Leishmaniasis, Toxoplasmosis, or Mycobacteriosis). In addition, the use of DNA oligonucleotides to direct a B cell immune response towards certain classes of immunoglobulins, thus bypassing and overcoming the adverse Ig-dependent diseases like Ig-E-mediated allergy is also an aspect of the present invention.

Use of DNA oligos to suppress immune reactions of the innate and acquired immune system is also an aspect of the present invention. DNA oligonucleotides can be used to suppress B- and T-cell responses towards transplantation antigens and thus induce transplantation tolerance. DNA oligonucleotides can further be used to suppress ongoing or manifested immune responses as it is the case during T- and B-cell dependent autoimmune diseases.

The pharmaceutical preparations of the present invention preferably comprise polynucleotides applied together with antigen either in free form or entrapped in liposomes. The science of forming liposomes is now well developed. Liposomes are unilamellar or multilamellar vesicles having a membrane portion formed of lipophilic material and an interior aqueous portion. The aqueous portion is used in the present invention to contain the antigen and, probably, protein material to be delivered to the antigen presenting cell. Conventional methods can be used to prepare liposomes. They are taken up by macrophages and dendritic cells in vivo and are thus particularly effective for delivery of antigen to these cells.

Liposomes may be manufactured by a rehydration entrapment method. Preferably the liposomes are prepared as follows. 18.0 mg phosphatidylcholine; 2.0 mg phosphatidylglycerol and 5.0 mg cholesterol, at a 2:0.2:1 ratio, are suspended in 5.0 ml chloroform in a 100 ml round bottom flask. The mixture is rotary evaporated under reduced pressure until a thin lipid film forms on the flask wall. Residual chloroform is removed by vacuum desiccation. 3 mg of ovalbumin is solubilized in 1.0 ml of PBS.

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This solution is slowly added to the dried lipid and hand-shaken until the lipids are resuspended. The crude protein liposome mixture is allowed to equilibrate for 30 min at room temperature, transferred to a microfuge tube and centrifuged at 6,000 rpm for 5 min in an Eppendorf microfuge. The mixture is then filter-extruded through a 0.2 µg Anotop10® syringe mount filter. To this mixture 10 nmol oligomer is added per 100 µl.

The pharmaceutical composition of the invention may advantageously be tested in mice. In such experiments mice were usually immunized by way of the hind footpad with 50 µl per foot of peptide liposome preparation. After four days, the draining popliteal lymph nodes (LN) were removed and a single-cell suspension was prepared. The cells were cultured for four days in the presence of IL-2 and a chromium release assay was performed utilizing the syngenic target cell EL-4 or the cell line EG-7 which is transfected with the gene for ovalbumin and thus presents ovalbumin peptides as antigen (Fig. 1). In some experiments EL-4 pulsed with the MHC class I (Kb) restricted ovalbumin peptide SIINFEKL was used as the target for kill.

The present invention also comprises methods of immunizing patients against a variety of diseases and conditions that have been referred to herein above or of treating patients suffering from one of the above referenced conditions or diseases. Formulations, routes of administration and doses have been identified herein above in connection with the discussion of the pharmaceutical compositions of the invention.

Finally, the invention relates to a method comprising

- (a) testing a nucleic acid molecule comprising a putative binding site of a transcription factor for toxicity;
- (b) modifying the nucleic acid sequence of said putative binding site comprised in said nucleic acid molecule which has proven toxic in step (a); and
- (c) repeating steps (a) and (b) one or more times until a non-toxic nucleic acid molecule has been identified.

Isolation of nucleic acid molecules to be tested, testing procedures as well as modification procedures for the nucleic acid sequences are well known in the art and had been described in the present specification, respectively.

An oligonucleotide that has been identified to be beneficial in accordance with the invention is IL-12p40 AGCTATGACGTTCCAAGG.

The figures show:

Figure 1: Cytolytic T cell induction in vivo with different ssDNA adjuvants.

Mice were injected with the antigen ovalbumin entrapped in liposomes using the indicated adjuvant. T cells were harvested from the mice and tested for the specific recognition of the antigen. The assay indicated a strong activation of the cellular immune system due to the adjuvant. AP-1, Stat-5/6, SIE, c/Myb, Stat3, Stat4, Stat5, SP-1, C/EBP, IL-13, Stat1, CRE, Erg and IRF-1 are binding sites for transcription factors.

Figure 2: A summary of cytolytic T cell assay.

Different sequences were tested. In vivo induced CTL were tested for specific antigen recognition. Data are expressed as lytic units.

A lytic unit is arbitrarily defined as the number of lymphocytes required to yield 30% specific lysis. The number is the lytic units per 10^6 effector cells. Lytic units are a way to compare cytolytic T cell populations.

Figure 3: Antibody production by ssDNA adjuvants post injection.

Endpoint antibody titer assay: Mice were injected with the antigen ovalbumin entrapped in liposomes using either no adjuvant or a TRE (transcription regulatory elements) adjuvant. The mice were boosted once.

The assay indicates a strong adjuvant effect for the enhancement of antibody production in response to injected antigen. The adjuvant-assisted increase in response is particularly strong for IgG2a and IgG2b. Of note is the differential induction of antibody isotype dependent on the adjuvant used indicating differential cytokine release.

Figure 4: Cytokine induction in vivo with transcription response element ssDNA adjuvant CRE.

The cytokine release pattern is induced by the transcription response element CRE. Mice were injected with the ssDNA and at the indicated time serum was sampled and cytokine release measured.

Figure 5: Phenotype changes (IL-2 receptor expression) of B and T cells in vivo with different transcription response element ssDNA adjuvants.

Analysis of a relevant cell surface marker after treatment with ssDNA. The IL-2 receptor binds and transduces a proliferation signal from IL-2 to cells of the immune system. The TRE sequences vary in their stimulatory capacity for inducing IL-2 receptor expression. Some TRE are inhibitory indicating a potential use for negative immunomodulation.

Figure 6: Tumor regression and control with ssDNA. ssDNA induces regression of prexistant tumor.

Mice were injected with tumorigenic numbers of a syngenic tumor cell. Four days after the challenge the mice were treated with ssDNA alone or ssDNA plus a subtumorigenic

number of tumor cells, which served as an antigen source. The progression of tumor growth was significantly controlled. Five mice were included per group.

Figure 7: Course of infection with *L. major* in ODN-treated mice.

BALB/c mice were injected with 2×10^6 *L. major* promastigotes into the right hind footpad. Oligonucleotide (ODN) (10 nmol) was given as treatment 2 h before and 10 h after infection. The mean percent increase (\pm SD of the footpad thickness is given (three mice per group). Closed symbols indicate the ODN-treated groups and the open circle the non-ODN-treated control group. ODN treatments are: closed circle, 1720; closed diamond, AP-1; closed triangle, 1668; closed square; IL-12-p40.

The examples illustrate the invention.

EXAMPLE 1

Bacterially derived sequences can be used as an adjuvant for cytolytic T cell activation in vivo.

Three sequences containing the sequence motif of 5'Pu-Pu-CpG-Py-Py-3' are described in the literature for having immunostimulating properties. One sequence is derived from the ampicillin resistance gene of *E. coli*, here termed AMP (TCATTGGAAAACGTTCTTCGGGGC). The second sequence is derived from a BCG gene and is termed BCG-A4A (ACCGATGACGTCGCCGGTGACGGCACACG). The third is a synthetic sequence claimed to be a prototype of bacterial CpG sequences, referred to by Krieg et.al. as 1668 (TCCATGACGTTCCTGATGCT). These sequences were synthesized to include a phosphorothioate linkage to reduce destruction by DNase. These oligomers served as an adjuvant in combination with ovalbumin to induce a cytolytic T cell response.

EXAMPLE 2

Bacterial CpG containing sequences may be toxic.

We observed that the above-described oligomers could be highly toxic in vivo for mice sensitive to TNF- α . The 1668 oligomer was previously described by Krieg and co-workers for its ability to induce murine B cell proliferation, induce IL-6 release from B cells and induce IFN- γ release from B cells and induce IFN- γ release from NK1.1 cells. We found in addition that 1668 and AMP were highly lethal in mice sensitized to the effects of TNF- α (Table 1). However, under certain circumstances these sequences may be useful therapeutically. One could foresee the use of these sequences once the risk is properly assessed.

Table 1 Death due to lethal shock

1668 + D-gal	5/5
1668 + LPS	3/3
AMP + LPS	2/3
Control	0/3

Ratio= mice killed/mice injected

For lethal shock, Balb/c mice were injected intraperitoneally with 10 nmol 1668 in 200 μ l PBS plus 20 mg D-galactosamine in 200 μ l PBS. Alternatively mice were injected intravenously with 10 nmol 1668, AMP or PBS followed at four hours with 50 μ g LPS.

EXAMPLE 3

Use of eukaryotic transcription regulatory elements or sequence manipulation prevents toxic shock symptoms.

Due to toxicity, the need is established for the discovery of non-toxic sequences for safe human and animal use. Since toxicity is at issue when developing vaccine adjuvants and therapeutics, we were interested to develop oligomers that circumvented toxicity but retained immunostimulatory properties. We screened eukaryotic sequences displaying the absence of lethality but maintaining immunostimulatory qualities. One such sequence was the cyclic AMP response element (CRE) which is the consensus binding site for the transcription factors CREB/ATF as well as the AP-1 family, sequence (GATTGCCTGACGTCAGAGAG) [Roesler, W. J. et al., J. Biol. Chem. 263, 9063-9066 (1988)]. Table 2 demonstrates the loss of lethality of the CRE sequence. To further evaluate the sequence specificity of these effects we made sequence exchanges between CRE and 1668. An exchange of only two nucleotides between CRE and 1668 resulted in a loss of lethality (Table 2).

Table 2 Sequences of oligomers and death due to lethal shock

a	
1668	TCCATGACGTTCTGATGCT
CRE	ATTGCCTGACGTCAGAGAGC
1668-CA	TCCATGACGTCAGTATGCT
CRE-TC	ATTGCCTGACGTTCTGAGAGC
b	
1668	5/5
CRE	0/5
1668-CA	0/3
CRE-TC	3/3

Lethality was determined as in example 2. The 1668 sequence fortuitously contains a combination of transcription response elements, namely the transcription factor binding sites (TGACGTTCC). This element represents the binding site for HSVIP04 (ATF), HSINS04 (CREB half site), CAMV35SR03 (HBP-1a yeast) or ADE422 (AP-1) in combination with an HSIL606 site which is a repressor site (sequence analysis from EMBL database Heidelberg). This sequence can be found in the 5' non-coding regions (promoters) of several eukaryotic cytokine genes including human IL-13 promoter and IL-12 p40 intron 1. The CRE sequence contains all the response elements cited above except for HSIL606 and it contains the full CRE palindromic sequence (TGACGTCA). In accordance with the invention, the CRE sequence did not induce death and changes in the 1668 eliminate toxicity.

TNF- α release is a hallmark of lethal toxic shock [Tracey, K. J. et al., Science 234, 470-474 (1986), Tracey, K. J. et al., Nature 330, 662-664 (1987)]. An exchange of only two nucleotides between CRE and 1668 resulted in a loss of macrophage induced TNF- α release activity. The sequence of the corresponding oligonucleotide is given in Table 2. The reported 6-mer active core sequence of 1668 contains the CpG flanked by two 5' purines and two 3' pyrimidines. The exchange of CA for TC does not affect this motif, however, TNF- α release was severely diminished. Thus, the broader core 8-mer sequence or the transcription response element and not the surrounding sequence environment was responsible for these effects. In accordance with the invention, when utilizing macrophage derived TNF- α release as a marker, the information comprised in the prior art 5'-Pu-Pu-CpG-Py-Py-3' motif alone was not satisfactory for predicting oligomer activity or toxicity. Additionally, in contrast to 1668, CRE did not induce IL-6 release in vivo or from the ANA-1 cell line in vitro.

EXAMPLE 4

ssDNA containing transcription response elements (TRE) serve as adjuvant for antibody production.

In accordance with the invention, eukaryotic transcription response elements relevant to the immune system serve as immune adjuvant. To test the adjuvant qualities of different sequences we injected mice with either free ovalbumin plus oligomer or liposome encapsulated ovalbumin plus oligomer. The mice were boosted at day 14 and after one week ovalbumin specific endpoint antibody titers were determined in an isotype specific ELISA.

Figure 3 shows that different sequences strongly potentiated the antibody response and induced class switching toward IgG1, IgG2a and IgG2b.

Liposomes containing ovalbumin were prepared as described above. For antibody induction, 300 µg ovalbumin in PBS or liposomes containing ovalbumin were injected +/- 10 nmol oligomer in the hind footpads of C57/B6 mice. A boost of the like inoculum was given after two weeks and one week later blood was extracted for serum antibody titering.

EXAMPLE 5

ssDNA containing transcription response elements serve as adjuvant for cellular immunity.

We have described the use of liposomes in combination with Quil A or QS-21 to induce cytolytic T cells (CTL) to either soluble antigen or peptides [Lipford, G. B., Wagner, H. & Heeg, K., Vaccine 12, 73-80 (1994), Lipford, G. B. et al., J. Immunol. 150, 1212-1222 (1993)]. Liposome entrapped antigen alone was an ineffective inducer of CTL activity, but with the addition of immunostimulatory saponins the inoculum became effective. To

test the in vivo T cell immunomodulatory potential of oligomers we utilized this vehicle to demonstrate primary activation of CTL. Figure 1 shows a substantial primary CTL response induced by an inoculum of ovalbumin liposomes plus ssDNA matching transcription response elements. The lytic units value interpolated from these curves was approximately 500 L.U. as compared to <20 L.U. for ovalbumin liposomes only (Table 3). CTL memory, an important quality for vaccine protection, could also be demonstrated with these inocula. If mice were rested for two weeks after the first injection and reinjected with the same inoculum, CRE recalled CTL displaying lytic units measured at approximately 1500 L.U. (Table 3). Additionally, when the inoculum was formulated with the immunodominant K^b restricted ovalbumin peptide SIINFEKL, the oligomers induced a specific primary CTL response. Thus, oligomers serve as a strong in vivo stimulus resulting in T cell activation and the proliferation of antigen specific CTL effectors. The inoculum can contain protein or peptide as the target antigen.

Table 3 Cytolytic T cell response induced by oligomer in
lytic units

	CRE	PBS
Primary CTL	526 L.U.	<20 L.U.
Secondary CTL	1555 L.U.	<20 L.U.

Several other sequences have been determined to have immunomodulatory effects. Table 4 list tested eukaryotic transcription response elements (TRE), which are preferably used in the present invention.

Table 4 Sequences of eukaryotic TRE tested

CRE	GATTGCCT <u>GACGTC</u> AGAGAG
IL-13	GGAATGACGTTCCCTGTG
AP-1	GCTTGATGACTCAGCCGGAA
SP1	TCGATCGGGGCGGGGCGAGC
C/EBP	TGCAGATTGCGCAATCTGCA
EGR	AGCGGGGGCGAGCGGGGGCG
GAS/ISRE	TACTTTCAGTTTCATATTACTCTA
SIE	GTCCATTTCCTCGTAAATCTT
STAT1	TATGCATATTCCTGTAAGTG
STAT3	GATCCTTCTGGGAATTCCTA
STAT4	CTGATTTCCCGAAATGATG
STAT5	AGATTTCTAGGAATTCATC
STAT5/6	GTATTTCCAGAAAAGGAAC
IRF-1	AAGCGAAAATGAAATTGACT
c-Myb	CAGGCATAACGGTTCGCTAG
NFkB	ATATAGGGGAAATTTCCAGC
HSINF	CAAAAAAATTTCCAGTCCTT
HSIL-6	ATGTTTTCCTGCGTTGCCAG
CRENFkB	CTCTGACGTCAGGGGAAATTTCCAGC

The relative strength of the various transcription response elements for adjuvant potential for CTL induction can be seen in fig. 2.

EXAMPLE 6

ssDNA containing transcription response elements induce cytokine release in vivo.

Mice were injected with formulations containing different oligomers plus liposome or liposomes containing 300 µg ovalbumin. Serum was collected at various times and analyzed for serum cytokine levels by specific ELISA.

In addition T cells produce IL-2 in response to ssDNA. It was found that cytokine release patterns are sequence dependent and thus the eukaryotic transcription response element used influences the cytokine release outcome and thus the biological effects. This aspect of the invention is highly relevant, because cytokine release patterns can be influenced by different transcription response elements. ssDNA can be used to induce cytokine release in vivo to produce a desired outcome. This outcome could be to produce immune enhancement or immune suppression.

ssDNA containing transcription response elements can break tolerance in T cells. It can be demonstrated that T cells induced to become tolerant (that is non-response to antigen signals) in vivo by SEB injection break tolerance if ssDNA is injected up to seven days post tolerance induction. This finding has relevance for the development of specific modulations for desired biological outcome. It can be foreseen that ssDNA can be used to elevate the immune response of immunocompromised patients.

EXAMPLE 7a:

ssDNA containing transcription response elements induce co-receptor and surface receptor change on B and T cells. This finding has relevance for the development of specific modulations for desired biological outcome.

Mouse spleen cells were harvested and cultured 24 hours in the presence of the list eukaryotic TRE. Cell surface markers were measured on T or B cell by FACS analysis.

Some of the transcription response elements have a positive effect and some have a negative effect. Both types of results are of potential medical use. If immune enhancement is desired a sequence inducing a given cell surface marker would be of use. If immune suppression is desired a sequence suppressing a given cell surface marker would be of use. Further guidance on such selection is provided in Example 7b.

EXAMPLE 7b:

ssDNA containing transcription response elements induce (a) expression of co-stimulatory molecules B7.1, B7.2 and CD40, (b) production of the cytokine IL-12, IL-6 and TNF- α on murine dendritic cells. These events (termed maturation and activation) are associated with the acquisition of professional antigen presenting activity to naive T cells.

Immature murine dendritic cells were grown from bone marrow cells in GM-CSF conditioned medium according to published protocols. At day 8 to 11 of culture, non-adherent cells were either MHC class II negative, or intermediate (termed immature DC) or high (termed mature DC). FACS[®] sorting of MHC class II intermediate of high cells revealed DC-like morphology. Bacterial DNA or ssDNA containing transcription response elements strongly upregulated CD80 (B7.1) CD86 (B7.2) CD40 and MHC class II molecules on immature DC, as measured by FACS analysis. In addition, i.s. (immune stimulating) DNA containing transcription response element triggered production of high concentrations of IL-12, TNF- α and IL-6. Finally, the i.s. DNA matured/activated DC (derived from sorted MHC class II intermediate cells) expressed professional APC function as assayed in an allogenic "mixed lymphocyte reaction" and in primary T cell cultures stimulated with the superantigen "staphylococcal enterotoxin B" (SEB). It is known that SEB does not require processing but requires professional

APC for presentation to naive V β 8 T cells. Conversion, as induced by ssDNA, of immature DC into professional APC explains the powerful adjuvant effect of ssDNA containing transcription response elements in *in vivo* on humoral and T cell response to poorly immunogenic antigens used for vaccination.

EXAMPLE 8

ssDNA containing transcription response elements can induce tumor control or regression.

Figure 6 demonstrates that mice challenged with tumor cells progress rapidly to display measurable subcutaneous tumors. These tumors are lethal. If ssDNA is injected four days post challenge the tumor regress or show retarded rates of growth. In addition if cells are provided as antigen the same observation is made.

EXAMPLE 9

Course of infection with *L. major* in ODN-treated mice.

BALB/c mice were injected with 2×10^6 *L. major* promastigotes into the right hind footpad. Oligonucleotide (ODN) (10 nmol) was given as treatment 2 h before and 10 h after infection. The mean percent increase (\pm SD of the footpad thickness is given (three mice per group). Closed symbols indicate the ODN-treated groups and the open circle the non-ODN-treated control group. ODN treatments are: closed circle, 1720; closed diamond, AP-1; closed triangle, 1668; closed square; IL-12-p40.

Claims

1. A pharmaceutical composition comprising
 - a) at least one fragment of a polynucleotide;
 - b) at least one antigen; and optionally
 - c) a pharmaceutically acceptable carrier and/or diluent.
2. The pharmaceutical composition according to claim 1 characterized in that the polynucleotide (a) comprises the sequence of a binding site for transcription factors or a part thereof or the sequence which is complementary to said binding site for transcription factors or a part thereof.
3. A pharmaceutical composition comprising
 - (a) a polynucleotide or an oligonucleotide comprising the sequence of a binding site for transcription factors or a part thereof or a polynucleotide or an oligonucleotide comprising a sequence which is complementary to said binding site for transcription factors or a part thereof; and optionally
 - (b) a pharmaceutically acceptable carrier and/or diluent.
4. The pharmaceutical composition according to any one of claims 1 to 3 characterized in that the polynucleotide is a DNA oligonucleotide.
5. The pharmaceutical composition according to claim 4 characterized in that the DNA oligonucleotide is single stranded.
6. The pharmaceutical composition according to any one of claims 1-5 characterized in that the polynucleotide comprises 5-40 nucleotides.

7. The pharmaceutical composition according to claim 6 characterized in that the polynucleotide comprises 15-25 nucleotides.
8. The pharmaceutical composition according to any one of claims 1-7 characterized in that the polynucleotide comprises the sequence
5'PuPuCGPyC
or a non-toxic derivative thereof.
9. A pharmaceutical composition according to any one of claims 1 to 9 wherein said binding site is or is derived from a eukaryotic binding site.
10. The pharmaceutical composition according to claim 9 wherein said eukaryotic binding site is a binding site for a cytokine.
11. The pharmaceutical composition according to any one of claims 2 to 10 wherein said part is a motif of a transcription factor binding site or a complementary sequence thereof.
12. The method of any one of claims 2 to 11 wherein said part comprises at least 7 nucleotides.
13. The pharmaceutical composition according to any one of claims 1 to 12 characterized in that the polynucleotide comprises at least one phosphorothioate linkage.
14. The pharmaceutical composition according to any one of the preceding claims 1 to 3, characterized in that it comprises a further adjuvant.
15. The pharmaceutical composition according to any one of claims 1 to 11, characterized in that the antigen (b) is selected from the group comprising peptides, polypeptides, proteins, polysaccharides, steroids and tumor cells.

16. The pharmaceutical composition according to any one of claims 1 to 15, characterized in that the composition is a vaccine.
17. The pharmaceutical composition according to claim 16 characterized in that the vaccine is used for the treatment of cancer.
18. The pharmaceutical composition according to claim 16 characterized in that the vaccine is used for the prophylaxis and/or treatment of pathogen microorganisms.
19. Use of an oligonucleotide or a polynucleotide as defined in any of the preceding claims for the preparation of a pharmaceutical composition for the modulation, enhancement or suppression of an immune response.
20. Use according to claim 19 wherein the modulation, suppression or enhancement is the result of a vaccination.
21. Use according to claim 19 wherein the modulation, suppression or enhancement is selected from the group break of tolerance, regulation of TH1/TH2 helper cell responses, switch of Ig classes, treatment of autoimmune responses and induction of tolerances.
22. Use of an oligonucleotide or a polynucleotide as defined in any of the preceding claims as an immune adjuvant.
23. A method for the identification of a nucleic acid sequence useful as an enhancer, modulator or suppressor of an immune response comprising
 - (a) testing a nucleic acid molecule comprising a putative binding site of a transcription factor for toxicity;

- (b) modifying the nucleic acid sequence of said putative binding site comprised in said nucleic acid molecule which has proven toxic in step (a); and
- (c) repeating steps (a) and (b) one or more times until a non-toxic nucleic acid molecule has been identified.

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TRE Adjuvant: In Vivo CTL Induction Specific Lysis of Ovalbumin Target Cells

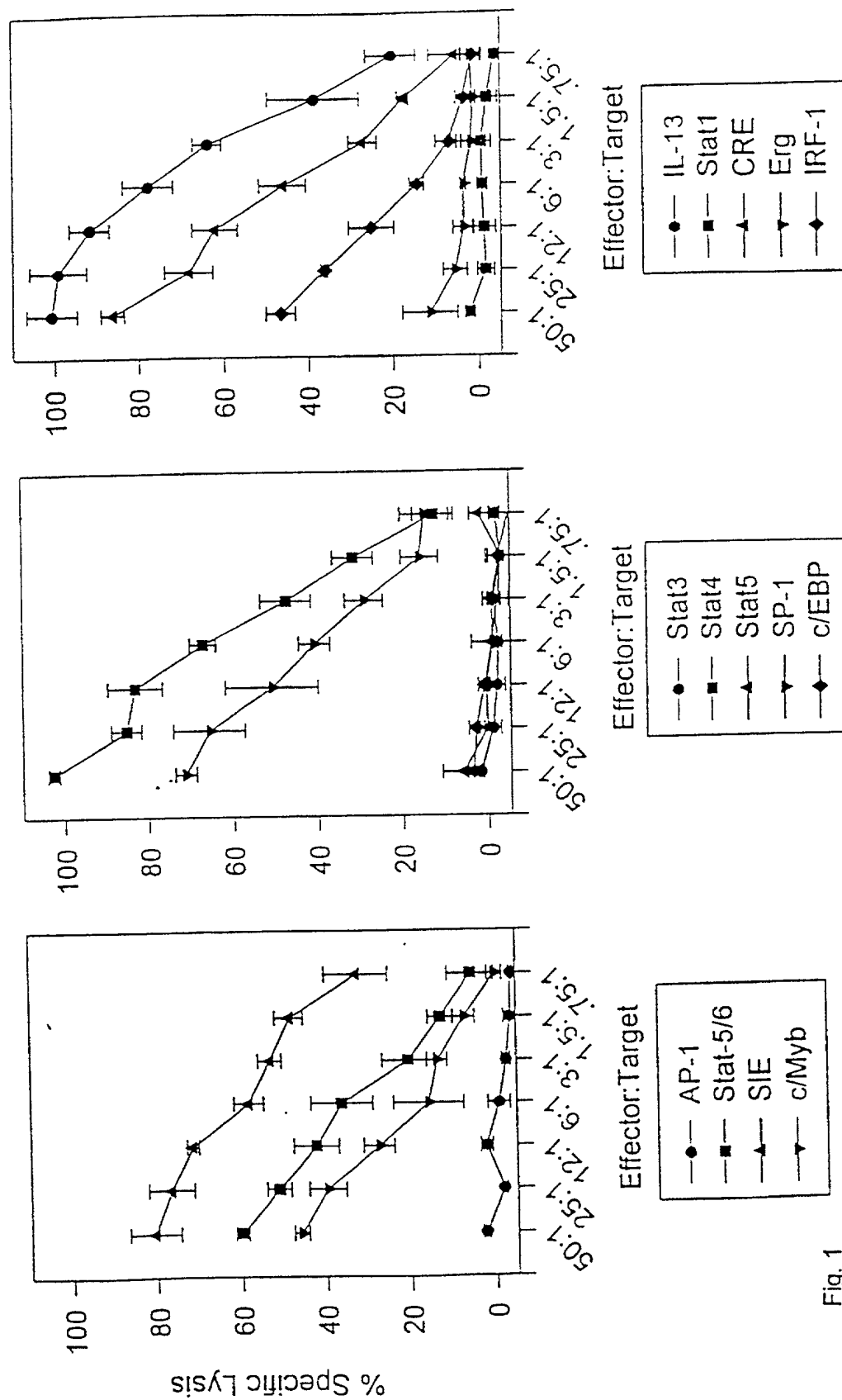


Fig. 1

TRE Adjuvant: In Vivo CTL Induction Specific Lysis in Lytic Units

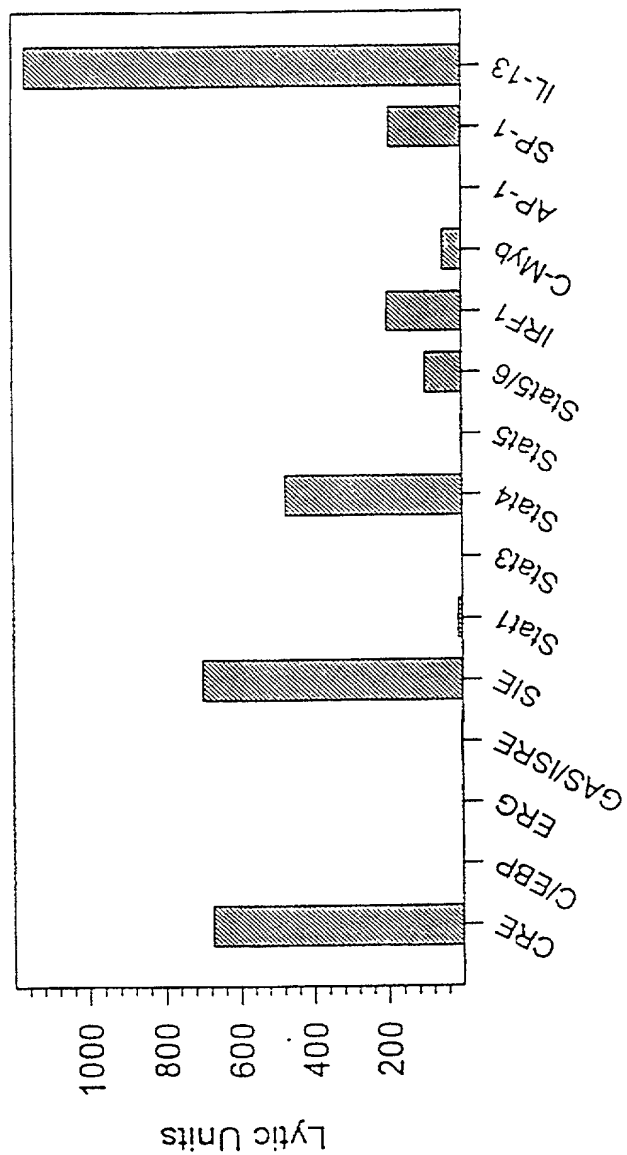


Fig. 2

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TRE Adjuvant: In Vivo Antibody Induction Specific Response to Antigen

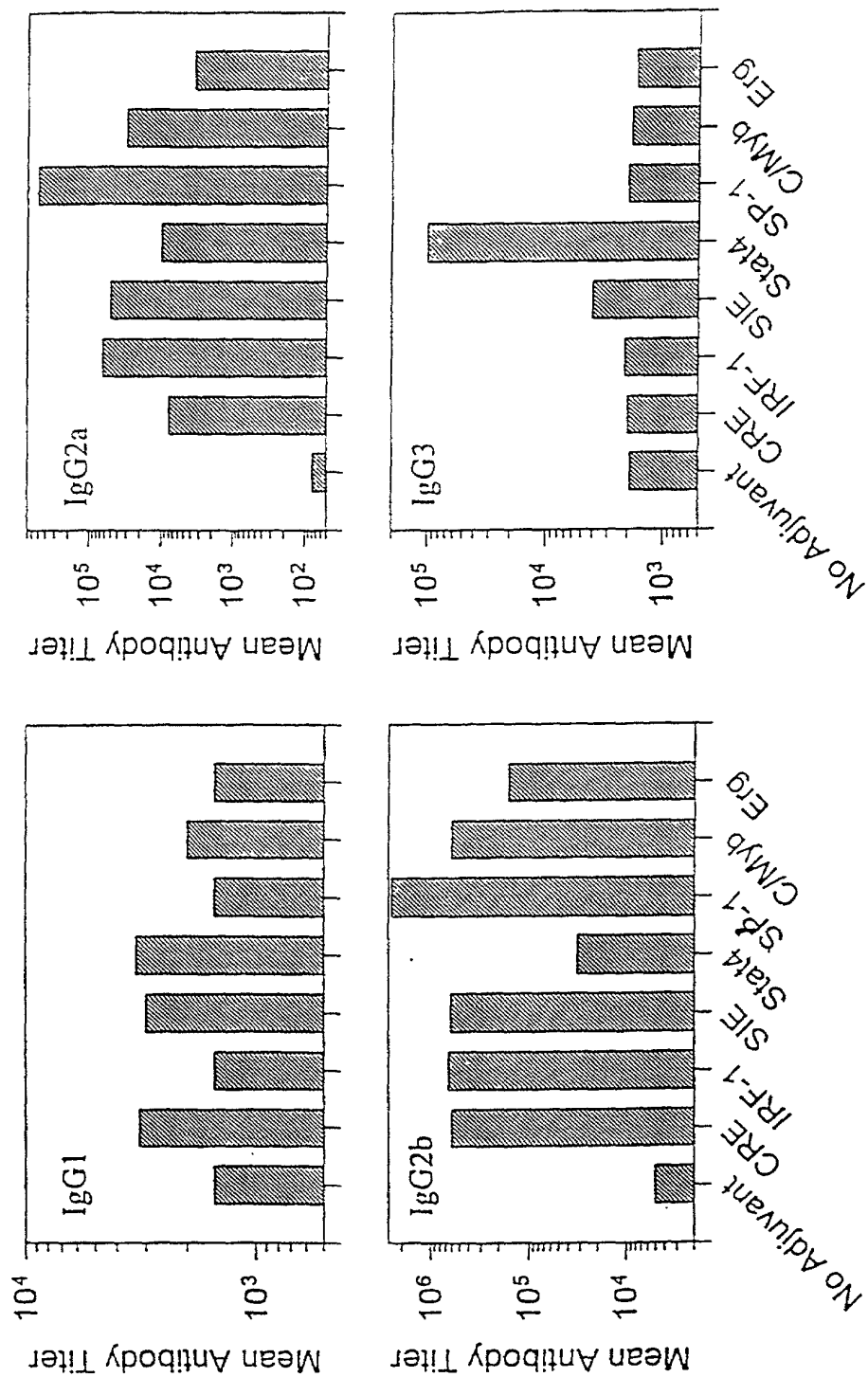


Fig. 3

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CRE: In Vivo Cytokine Induction

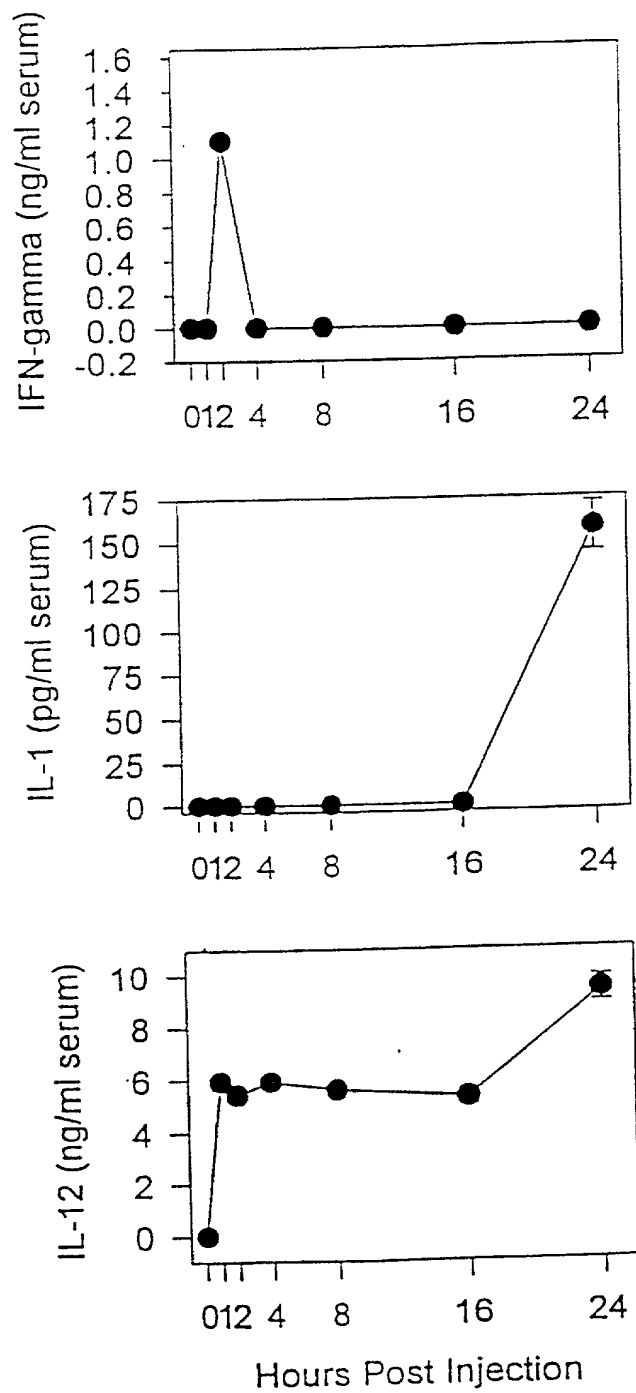


Fig. 4

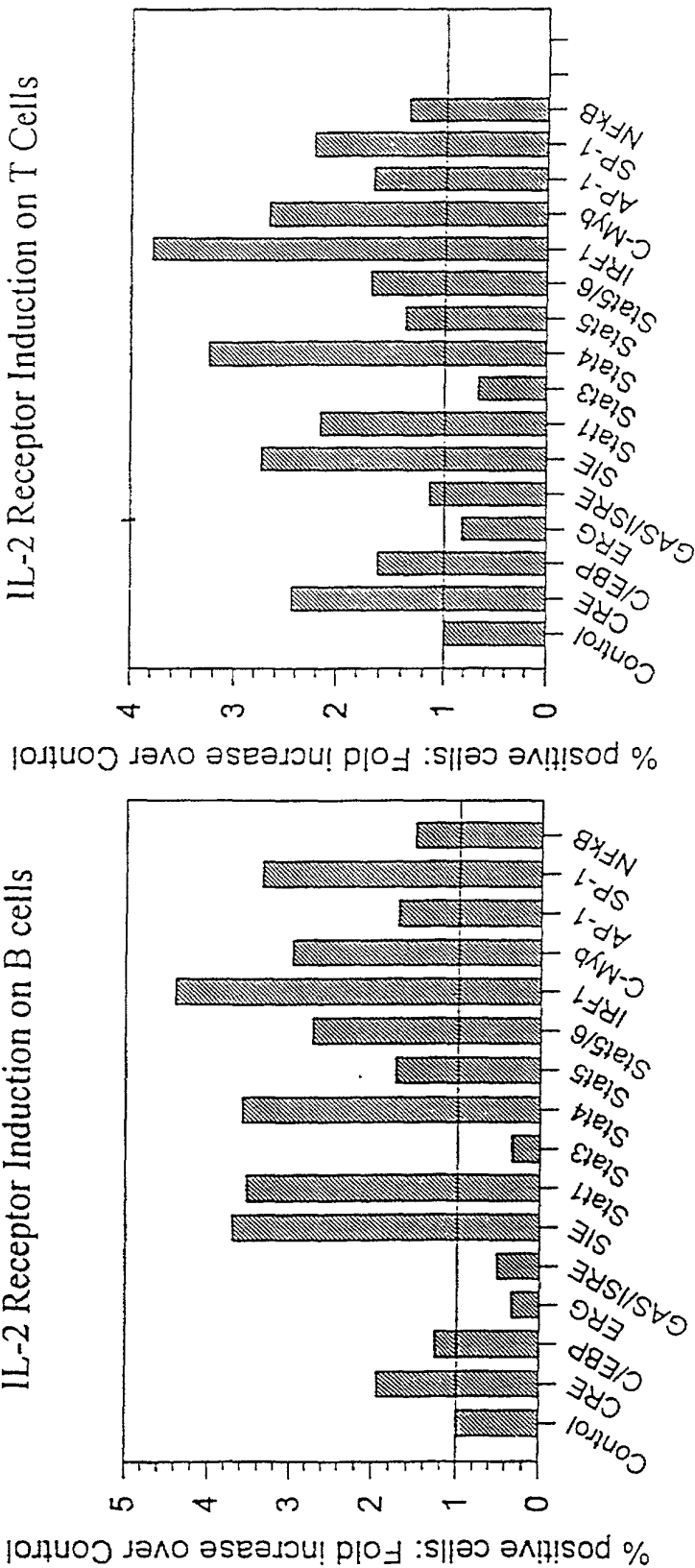


Fig. 5

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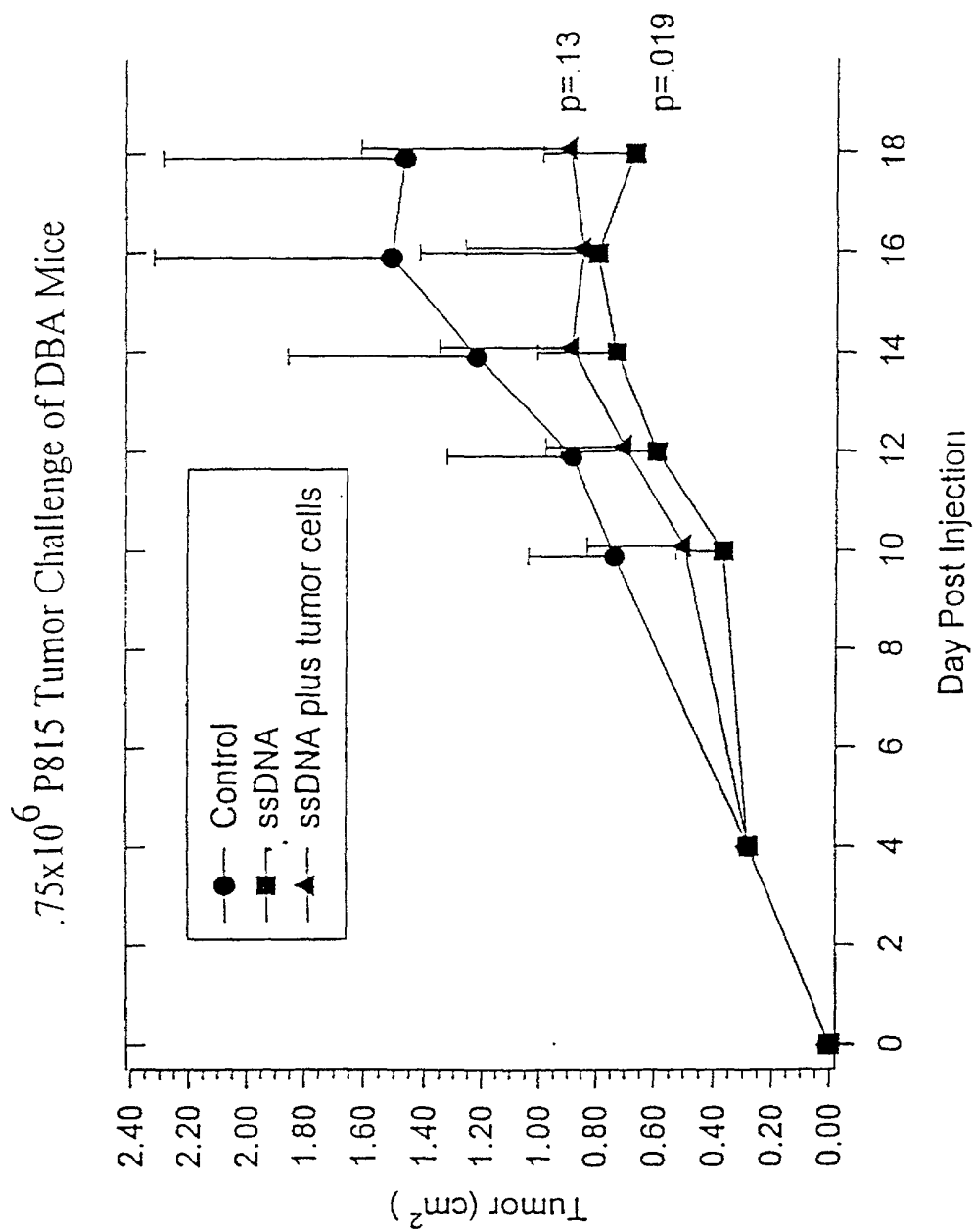


Fig. 6

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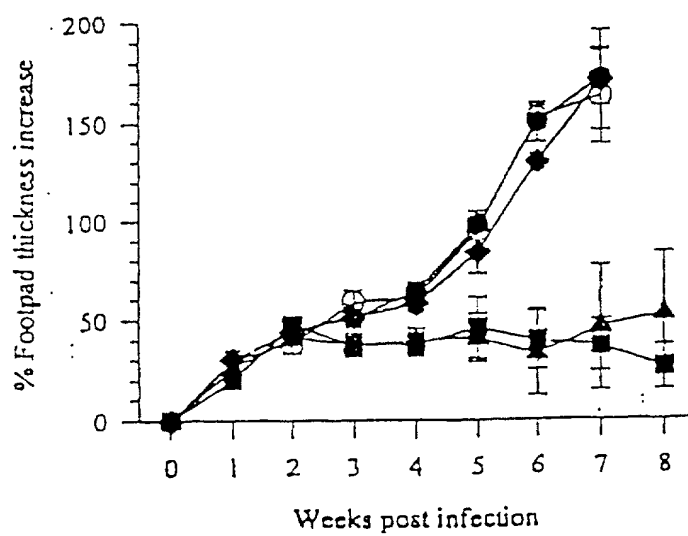


Fig. 7



DECLARATION FOR PATENT APPLICATION

a below named inventor, I hereby declare that:

My residence, post office address and citizenship are as stated below next to my name.

I believe I am the original, first and sole inventor (if only one name is listed below) or an original, first and joint inventor (if plural names are listed below) of the subject matter which is claimed and for which a patent is sought on the invention entitled

**PHARMACEUTICAL COMPOSITIONS COMPRISING A POLYNUCLEOTIDE AND
OPTIONALLY AN ANTIGEN ESPECIALLY FOR VACCINATION**

the specification of which is attached hereto unless the following is checked:

☒ [X] was filed on January 23, 1998 as United States Application No. 09/355,254.

I hereby state that I have reviewed and understand the contents of the above identified specification, including the claims, as amended by any amendment referred to above.

I acknowledge the duty to disclose information which is material to the examination of this application in accordance with Title 37, Code of Federal Regulations, §1.56.

I hereby claim foreign priority benefits under Title 35, United States Code, §119(a)-(d) or §365(b) of any foreign application(s) for patent or inventor's certificate, or section 365(a) of any PCT International application designating at least one country other than the United States listed below and have also identified below any foreign application for patent or inventor's certificate or PCT International application having a filing date before that of the application on which priority is claimed:

Prior Foreign PCT International Application(s) and any priority claims under 35 U.S.C. §§119 and 365(a), (b):

PCT/EP98/00367	23/01/98	Priority Claimed
(Number)	(Country-if PCT, so indicate) (DD/MM/YY Filed)	<input checked="" type="checkbox"/> [X] <input type="checkbox"/> []
97101019.4	EP 23/01/97	YES NO
(Number)	(Country) (DD/MM/YY Filed)	<input checked="" type="checkbox"/> [X] <input type="checkbox"/> []
		YES NO
(Number)	(Country) (DD/MM/YY Filed)	<input type="checkbox"/> [] <input type="checkbox"/> []
		YES NO

I hereby claim the benefit under Title 35, United States Code, §119(e) of any United States provisional application(s) listed below:

(Application Number) (filing date)

(Application Number) (filing date)

I hereby claim the benefit under Title 35, United States Code, §120 of any United States application(s), or §365(c) of any PCT International application(s) designating the United States of America listed below and, insofar as the subject matter of each of the claims of this application is not disclosed in the prior United States or PCT International application in the manner provided by the first paragraph of Title 35, United States Code, §112, I acknowledge the duty to disclose material information as defined in Title 37, Code of Federal Regulations, §1.56 which became available between the filing date of the prior application and the national or PCT International filing date of this application:

(Application No.) (filing date) (status-patented, pending, abandoned)

(Application No.) (filing date) (status-patented, pending, abandoned)

PCT International Applications designating the United States:

(PCT Appl. No.) (U.S. Ser. No.) (PCT filing date) (status-patented, pending, abandoned)

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